

Original Article

Fungi diversity assessment under different tree species of Shere Hills soils in Jos North Local Government Area Plateau State, North Nigeria

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ABSTRACT

This study assessed the fungi composition under three different forest tree species cover and bare land in Shere Hills Jos, North Nigeria. Plants species considered are *Dichrostachys*, *Terminalia*, and *Bombax*. Two meters by 2 m plot under forest cover and adjacent open area were set up. Fungi species diversity indices (Shannon-Wiener index and species evenness) were calculated. Results revealed that there were 49 species distributed into 28 families (293 individuals) under the tree cover, while there were 53 species distributed into 33 families (451 individuals) in the open vegetation. The Jaccard index of similarity (79.49%) was high between the study sites. The result of this study showed that forest cover has a beneficial effect on soil properties and nutrient pool. Plants species richness and microbes population were also higher under the forest cover than in the open area. Species diversity and richness, soil nutrient, and microbial population were greatly influenced by forest cover.

Keywords: Soils, fungi, Shere Hills, diversity, tree species, plate count

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INTRODUCTION

The forest canopy is defined as “the top layer of a forest or wooded ecosystem consisting of overlapping leaves and branches of trees, shrubs, or both.”^[1] Forest canopy is also frequently explained as the percentage area occupied by the vertical projection of tree crowns; a proportion of the forest floor covered by the vertical projection of the tree crowns.^[2] The tree canopy is very important in that it modulates the availability and variability of some key resources for the organisms living in the understorey, thus affecting its own regeneration.^[3] Plant species can influence the composition of underlying soil microbial communities.^[4] These influences can be due to differences in forest cover, rooting depth, and litter quality and quantity^[5] or to secondary effects on soil pH, moisture, and nutrient levels. Microbes (bacteria, archaea, fungi, and protozoan) are very important in all processes related to soil function.^[6] The microbial constituents of soil

are entirely responsible for the breakdown of organic matter and the degradation of toxic molecules.^[7] Overall, denser forest cover implies higher rate of respiration.^[8]

Forest canopies contain a major portion of the diversity of organisms on earth and constitute the bulk of photosynthetic active foliage and biomass in forest ecosystems.^[1] Soils are highly diverse habitats that mediate biogeochemical processes of global importance, yet our understanding of how these functions are influenced by microbial biodiversity is only slowly advancing. At present, microbial community composition is an important determinant of ecosystem process rates and identifying microbial community composition has become an essential component for predicting ecosystem responses to environmental change. Furthermore, it is essential to simultaneously study the bacterial and fungal communities in soil to understand microbial influences on ecosystem processes, because these communities mediate different ecological functions in soil.^[8]

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This study was carried out in order to provide information on fungi diversity in soils under tree canopy of major tree species in Shere Hills. The specific objectives of this study are to produce a checklist of fungi in the various soil samples and compare the fungi diversity across different tree species and plots on the field.

MATERIALS AND METHODS

Study Site

Shere Hills are located in Jos, Plateau situated about 10 km to the East of Jos metropolis the capital of Plateau State in the middle belt region of Nigeria with a latitude of 9°57'N 9°03'E and longitude 9.950°N 9.050°E and have an elevation of 1829 m (6001 ft) above sea level, the Shere Hills are the highest point of the Jos Plateau and they form the third highest point in Nigeria after the Chappal Waddi on the Mambilla Plateau averaging about 2419 m or 7936 feet above sea level and mount Dimlang (Vogel peak) on the Shebshi mountains reaching a height of about 2042 m or 6699 feet above sea level.^[9]

Sampling Procedures

The study was conducted in Shere Hills, Jos North, Plateau State, Nigeria. Three tree species covers were selected within the Shere Hills forest. Two meters by 2 m plot were marked out within each of the three tree cover using a measuring tape. Three other points (open area) were selected adjacent to each of the tree cover at a distance of 5 m away from the tree canopy (open area), where 2 m by 2 m plot was also marked out. This study was carried out between August and September 2020, the peak of the raining season because it is expected that the plant growth will be at its peak. The species that form the forest cover used in this study were *Dichrostachys*, *Terminalia*, and *Bombax*.

Determination of the Fungi Population

The assessment of the soil sample for fungi count was determined by standard pour plate method as described by Nwachukwu and Akpata (2003).^[10] Culturing, inoculum from serial dilution of the same soil suspension, was transferred into sterile Petri dishes followed by sterile molten Sabouraud Dextrose agar of 40–42°C, mixed, allowed to solidified, and incubated in an incubator at 25°C for 5 days. Growth of microorganism was observed after incubation period and the population determined by standard microbiological method putting into consideration the dilution factor.^[10]

Data and Statistical Analysis

A number of fungi species (i.e., species richness), genera, and families were established for each plot. The Simpson index of diversity (1-D) was also calculated

$$D = \frac{\sum(n-1)}{N(N-1)}$$

Where, N is the total number of individual species present at the study sites and n is the number of species present at the study sites. Fungi species diversity index H' of each sample plot was calculated using the method prescribed by.^[11]

$$H' = -\sum P_i \ln P_i$$

Where, $P_i = n_i/N$ is the relative abundance proportion of i species and $\ln =$ Natural logarithm N is the total number of individuals present at the study sites. n_i is the number of species present at the study sites

The degree of similarity in species composition between open and cover area was compared using Jaccard index of similarity (J):

$$J = \frac{j}{r} \times 100$$

Where, j is the number of species found in both cover and open plots and r is the number of species found in each of the plots added together except common species in the two sites (i.e., number of species found in only one or the other).

The evenness index^[12] was calculated thus:

$$E = \frac{H'}{\ln S}$$

Where, S is the total species number in each site H' is the diversity index and $\ln =$ natural logarithm.

RESULTS AND DISCUSSION

Fungi Encountered and Abundance in the Study Area

The details of the differences in the number of fungi found under individual plant species of focus and on the bare land in Shere Hills are presented in Table 1. The study revealed that the following species *Sporangium* (159), *Paecilomyces* (27), *Hirsutella* (22), *Entomophage* (29), and *Ascosphaera aggregate* (16) were dominant in the open area (Bare land), while *Acremonium* (28), *Sporangium* (20), *Phoma* (25), *Penicillium* (16), *Aspergillus Alternaria conidia* (28), *Aspergillus conidia* (88), *Verticillium lecanii* (33), *Sporangium* (63), *Tolypocladium* (28), *Penicillium* (35), *Torribiella* (55), *Hirsutella* (35), *Metarhizium anisopliae* (28), yeast *Psuedohyphae* (17), *Penicillium* (51), *Aspergillus* (56), *Sporangium* (37), and *Cladosporium conidia* (17) are the dominant fungi in the area underneath the forest cover [Table 1]. A total number of 1032 fungi count were recorded with the highest fungi count of 400 observed under *Terminalia* tree canopy, followed by *Bombax* and bare land recorded the least number of fungi across the study area.

Diversity and Evenness of Fungi Colony

The result on diversity and evenness of fungi colony as revealed in Table 2 shows that plot four had the highest value for fungi diversity ($H = 1.42$) for *Dichrostachys*. This was followed by plot five with diversity of 1.24, plot one ($H = 1.12$), and plot two ($H = 0.88$) while plot three was the least with a diversity of 0.86. The highest value of diversity of fungi colony for *Terminalia* was seen in plot four ($H = 1.87$), this was followed by plot three with diversity of 1.61, plot two ($H = 1.47$), and plot one (1.22) while plot five was also the least with a diversity of 0.64. The highest value for fungi diversity on *Bombax* was seen in plot one ($H = 1.54$), this was followed by plot five with diversity of 1.37, plot two ($H = 1.09$), and plot three (0.82) while plot four was also the least with a diversity of 0.00. The colony of fungi diversity found on bare land shows highest value in plot five with a diversity of ($H = 1.66$), this was followed by plot one with diversity of 1.25, plot four ($H = 0.79$), and plot two (0.62) while plot three was also the least with a diversity of 0.00.

The result on fungi colony evenness for *Dichrostachys* shows highest value ($J' = 0.89$) in plot five. This was closely

followed by plot four ($J' = 0.88$), plot two ($J' = 0.80$), and plot three ($J' = 0.78$) and plot one was the least with $J' = 0.63$. On *Terminalia*, evenness of fungi value was highest ($J' = 0.96$) in plot four, this was followed by plot five ($J' = 0.92$), plot two ($J' = 0.91$), plot three ($J' = 0.90$), and the least on plot one with value of $J' = 0.88$. The result on species evenness for *Bombax* shows highest value ($J' = 0.99$) in plot two, this was followed by plot one ($J' = 0.96$), plot five ($J' = 0.85$), plot three ($J' = 0.75$), and the least on plot four with least value of $J' = 0.00$ while the colony of fungi found on bare land shows highest value in plot five with evenness value of $J' = 0.99$, this was followed by plot one and two ($J' = 0.90$), respectively, plot four ($J' = 0.72$), and the least on plot three with least value of $J' = 0.00$ [Table 2].

The study shows that there was no significant difference between the fungi diversities across the different plots ($P \leq 0.05$) [Table 3]. The Fungi diversities under *Dichrostachys*, *Terminalia*, *Bombax* and on bare ground were not significantly different ($P \leq 0.05$) [Table 4] with means values as shown in Figure 1.

DISCUSSION

Soil microbial communities are an integral component of many ecosystem processes.^[13] The microbial population of the area underneath the tree cover was found far higher than those of the area outside tree cover. Woody plant canopies alter the microenvironment and physical and fertility conditions of soil.^[14] Trees modify microenvironment in terms of reduced soil and air temperatures, wind speed, and irradiation, resulting into reduced soil water evaporation and increased relative humidity.^[15] It has also been pointed out that trees also acquire nutrients from deeper soil layers and redistribute them at the surface through litter fall which enhances soil carbon and nutrients. All these may create a favorable condition that benefits microbial activity^[16] have also reported that soil water and temperature were influenced by tree cover litter accumulated and this creates a favorable condition that enhances microbial activity. Most microorganisms (such as fungi) thrive best in a narrow pH range near neutrality, between pH 6.5 and 7.5; very few thrives below pH 4.0.

The relative abundance of the fungi encountered in the areas underneath cover is indication that soils underneath forest

Table 1: Comparison of fungi composition and number encountered across the study sites.

| Plants | Colony |
|----------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <i>Dichrostachys</i> | <i>Acremonium</i> (28), <i>Verticillium</i> (8), <i>Paecilomyces</i> (2), <i>Rhizopus</i> (14), <i>Phoma</i> (25), <i>Penicillium</i> (16), <i>Aspergillus</i> (8), <i>Trichoderma</i> (8). |
| <i>Terminalia</i> | <i>Alternaria conidia</i> (28), <i>Aspergillus conidia</i> (88), <i>Verticillium lecanii</i> (33), <i>Tolypocladium</i> (28), <i>Penicillium</i> (35), <i>Torrubiella</i> (55), <i>Hirsutella</i> (35), <i>Metarhizium anisopliae</i> (28), <i>Ascospaera aggregate</i> (7) |
| <i>Bombax</i> | Yeast <i>psuedohyphae</i> (17), <i>Penicillium</i> (51), <i>Aspergillus</i> (56), <i>Rhizopus</i> (15), <i>Alternaria conidia</i> (14), <i>Blastospores</i> (1), <i>Cladosporium conidia</i> (17), <i>Histoplasma capsulate</i> (8) |
| Bare Land | <i>Paecilomyces</i> (27), <i>Hirsutella</i> (22), <i>Pandora</i> (7), <i>Entomophage</i> (29), <i>Torrubiella</i> (14), <i>Aspergillus</i> (4), <i>Ascospaera aggregate</i> (16), <i>Tolypocladium inflatum</i> (9). |

Source: Fieldwork, 2020

Table 2: Summary characteristics of fungi diversity among different plots

| Plants | Plot 1 | | Plot 2 | | Plot 3 | | Plot 4 | | Plot 5 | |
|----------------------|--------------------------|--------------------|--------------------------|--------------------|--------------------------|--------------------|--------------------------|--------------------|--------------------------|--------------------|
| | Shannon wiener index (H) | Evenness index (%) | Shannon wiener index (H) | Evenness index (%) | Shannon wiener index (H) | Evenness index (%) | Shannon wiener index (H) | Evenness index (%) | Shannon wiener index (H) | Evenness index (%) |
| <i>Dichrostachys</i> | 1.12 | 0.63 | 0.88 | 0.80 | 0.86 | 0.78 | 1.42 | 0.88 | 1.24 | 0.89 |
| <i>Terminalia</i> | 1.22 | 0.88 | 1.47 | 0.91 | 1.61 | 0.90 | 1.87 | 0.96 | 0.64 | 0.92 |
| <i>Bombax</i> | 1.54 | 0.96 | 1.09 | 0.99 | 0.82 | 0.75 | 0.00 | 0.00 | 1.37 | 0.85 |
| Bare land | 1.25 | 0.90 | 0.62 | 0.90 | 0.00 | 0.00 | 0.79 | 0.72 | 1.66 | 0.93 |

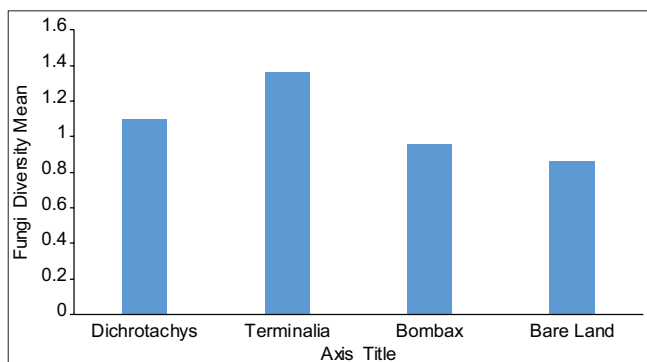
Table 3: Analysis of variance of fungi diversity across different plots

| Source | Sum of squares | df | Mean square | F | Sig. |
|----------|----------------|----|-------------|-------|---------------------|
| Variable | 0.700 | 3 | 0.233 | 0.898 | 0.464 ^{NS} |
| Error | 4.158 | 16 | 0.260 | | |
| Total | 4.858 | 19 | | | |

WHERE: NS: Not significant

Table 4: Standard deviation±mean effects of fungi diversity across different plots

| Variables | Fungi diversity index |
|----------------------|------------------------|
| <i>Dichrostachys</i> | 0.24±1.10 ^a |
| <i>Terminalia</i> | 0.47±1.36 ^a |
| <i>Bombax</i> | 0.60±0.96 ^a |
| Bare land | 0.63±0.86 ^a |
| SE | 0.23 |

Means on the same column with the same letter superscript do not differ significantly from each other ($P=0.0$)**Figure 1:** Mean effects of fungi diversity across different plots

cover is rich in biodegradable organic matters and these are very important in humus formation. This might have accounted for the usual fertile land underneath tree cover. In addition, excretions from soil microorganisms affect water and air movement within the soil. In the course of this study, it was observed that worm casts are more common in the areas underneath tree cover compared to the open area. This is an indication of the higher breaking down of organic matter underneath the tree cover^[17] reported that some fungi function largely in the breakdown of complex organic molecules like lignin (a compound that is resistant to bacteria degradation).

CONCLUSION

The study revealed that the number of fungi recorded under the tree cover was higher than those found in the open area. *Terminalia* tress also recorded the highest number of fungi when compared with other tree canopies and the bare land. The families fungi most abundantly represented in the area

underneath the forest cover are *Acremonium*, *Phoma*, *A. conidia*, *A. conidia*, *V. lecanii*, *Tolypocladium*, *Penicillium*, and *M. anisopliae* while the families most abundantly represented in the open areas are *Paecilomyces* and *Entomophage*. The Jaccard index of similarity indicated high similarity in richness, composition, and relative abundance between the two study areas. It is, therefore, suggested that strong regulatory measures should be put in place and also effort to reestablish the depleted tree species populations should be encouraged. Disturbance, especially through human activities, should be discouraged or completely avoided because of their negative impacts on plant species diversity, nutrients availability, and microbial population.

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CONFLICTS OF INTEREST

There are no conflicts of interest during this study.

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